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# TWO NOVEL STEROIDS FROM EUPHORBIA OFFICINARUM LATEX

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Two novel steroids,  $3\beta$ , $7\alpha$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -cholest-8-en-11-one (2) and  $3\beta$ , $7\beta$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -cholest-8-en-11-one (3) were isolated from the latex of *Euphorbia officinarum*. Their structures were established on the basis of NMR and MS studies.

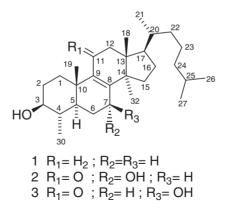
# **INTRODUCTION**

The latex of the Moroccan endemic plant *Euphorbia officinarum* L. (Euphorbiaceae) has been used in folk medicine to treat various skin and ophthalmologic diseases in low concentration due to its high toxicity [1]. Characterization of its components is necessary to fully develop its therapeutic potential.

Nine compounds with triterpenic and steroideal skeleton have been previously isolated [2] from the methanolic extract of the latex of this plant, which were identified as: lupeol, lupeol acetate, lanostenol, lanosterol, 24-methylene lanostenol,  $4\alpha,14\alpha$ -dimethyl-24-methylen-5 $\alpha$ -cholest-8-en-3 $\beta$ -ol,  $4\alpha,14\alpha,24(R)$ -trimethyl-5 $\alpha$ -cholesta-8,25(27)-dien-3 $\beta$ -ol,  $4\alpha,14\alpha$ -dimethyl-5 $\alpha$ -cholesta-8,24-dien-3 $\beta$ -ol and  $4\alpha,14\alpha$ -dimethyl-5 $\alpha$ -cholesta-8,24-dien-3 $\beta$ -ol and  $4\alpha,14\alpha$ -dimethyl-5 $\alpha$ -cholesta-8,26-ol (1). Further examination of this extract led to the isolation of two new steroids (2 and 3). This article deals with their structural elucidation.

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#### **RESULTS AND DISCUSSION**

Both compounds (2 and 3) were obtained as white needles and they were assigned the molecular formula  $C_{29}H_{48}O_3$  (HREIMS). The IR spectra of both compounds showed absorption bands for a  $\gamma$ -hydroxy- $\alpha,\beta$ -unsaturated ketone ( $\nu_{max}$  3474, 1642 cm<sup>-1</sup>). The <sup>1</sup>H NMR and <sup>13</sup>C NMR of both compounds showed the presence of three tertiary and two secondary methyl groups, an isopropyl group [2:  $\delta_{\rm H}$  0.86, 0.87 (each 3H, d); 3:  $\delta_{\rm H}$  0.87, 0.88 (each 3H, d)], nine methylene groups, five methine groups, two hydroxymethine groups [2:  $\delta_{\rm H}$  3.17 (1H, ddd), 4.35 (1H, dd);  $\delta_{\rm C}$  76.3 (d), 67.5 (d); 3:  $\delta_{\rm H}$  3.07 (1H, ddd), 4.41 (1H, dd);  $\delta_{\rm C}$  76.0 (d), 68.7 (d)], a tetrasubstituted double bond [2:  $\delta_{\rm C}$  160.5 (s), 140.6 (s); **3**:  $\delta_{\rm C}$  160.0 (s), 141.9 (s)], and a conjugated ketone [**2**:  $\delta_{\rm C}$  200.9 (s); **3**: 200.8 (s)]. A significative difference between these compounds lies in the coupling constants observed for one of the methine protons, assigned by 2D-NMR spectroscopy as H-7 [2:  $\delta_{\rm H}$  4.35 (1H, dd, J = 4.8, 1.1 Hz); 3:  $\delta_{\rm H}$  4.41 (1H, dd, J = 8.9, 8.9 Hz)]. A molecular model analysis shows that a difference in the configuration of the hydroxyl group at C-7 justifies the observed coupling constants. In compound 2, an  $\alpha$  configuration for the hydroxyl group at C-7 implies a half chair configuration for ring B and leaves H-7 $\beta$  bisecting both H-6 $\alpha$  and H-6 $\beta$  yielding small coupling constants; on the other hand, a  $\beta$  configuration for the hydroxyl group at C-7 leaves H-7 $\beta$  in *anti* and gauge relationships to H-6 $\beta$  and H-6 $\alpha$ , respectively, causing larger coupling constants [3]. These assignations are supported by the HMBC and NOESY spectra of compound 2. The NOESY data (Fig. 1) exhibited cross correlations for H-3 $\alpha$  (with H-5 $\alpha$ ), H<sub>3</sub>-18 (with H-20),  $H_3$ -32 (with H-12 $\alpha$  and H-17 $\alpha$ ), and H-7 $\beta$  (with H-15, H-15', H-6, H-6'). This analysis allows the assignment for **2** as  $3\beta$ , $7\alpha$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ ergost-8-en-11-one and for **3** as  $3\beta$ , $7\beta$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -ergost-8-en-11one. Further support for these assignments follows from the comparison of the <sup>1</sup>H NMR and <sup>13</sup>C NMR data of compound **2** with those of  $3\beta$ ,  $7\alpha$ -dihydroxy- $4\alpha$ ,  $14\alpha$ dimethyl-5 $\alpha$ -ergosta-8,24(28)-dien-11-one [4] and the comparison of the <sup>1</sup>HNMR and  ${}^{13}CNMR$  data of compound 3 with those of lucidenic acid N [5], which has an identical structure of the ring system with that of compound 3.

Both lucidenic acid N and  $3\beta$ , $7\alpha$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -ergosta-8,24(28)dien-11-one have cytotoxic [5] and antitumor-promoting [4] activities, respectively, what suggest compounds **2** and **3** to be potentially bioactive.

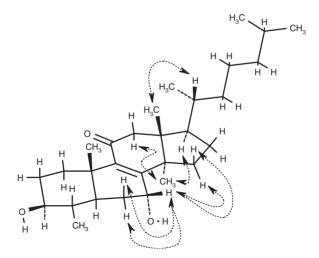


FIGURE 1 Selected NOESY correlations observed for compound 2.

# EXPERIMENTAL SECTION

#### **General Experimental Procedures**

Melting points were measured with a Reichert-Jung Kofler block and are uncorrected. Optical rotations were determined with a Perkin-Elmer 241 polarimeter. IR spectra were recorded on a Perkin-Elmer 881 spectrophotometer. <sup>1</sup>H and <sup>13</sup>C NMR measurements were obtained on a Varian INOVA 400 NMR spectrometer with SiMe<sub>4</sub> as internal reference. Mass spectra were recorded on VG Platform II and VG Autospec spectrometers at 70 eV. HPLC was performed with a Hitachi/Merck L-6270 apparatus equipped with a UV-Vis detector (L 4250) and a differential refractometer detector (RI-71). TLC was performed on 0.2 mm Merck Kiesegel 60 F<sub>254</sub>, plates. Silica gel (Merck) was used for column chromatography. Purification by HPLC was accomplished using a silica gel column (LiChrospher<sup>®</sup> Si 60, 10 µm, 1 × 25 cm long).

## **Plant Material**

Latex from *E. officinarum* was collected in May 1999 from plants of the North Atlantic coast of Agadir, Morocco. Latex was obtained by making repeated cuts along the stems of the plant, using a knife, and collecting the white milky exudates. A voucher specimen is deposited at the Herbarium of the laboratoire de Chimie des Substances Naturelles et Hétérocycles, Université Cadi Ayyad.

# **Extraction and Isolation**

The latex (0.5 L) from *E. officinarum* was allowed to dry and the resulting coagulum (200 g) was extracted with MeOH (1.5 L), employing a Sohxlet apparatus. The MeOH solution was cooled to yield a precipitate (60 g) which was subjected to silica gel column chromatography. Elution of the column with hexane:ethyl acetate (95:5) afforded three fractions [fraction (weight after solvent removal)]: A (5 mg), B (4 g), C (53 g) and elution with hexane:ethyl acetate (90:10) yielded a fourth fraction

[D (16 mg)]. Analysis of fractions A–C by chromatography on silica gel with 10% AgNO<sub>3</sub> yielded already known compounds [2]. Purification of the more polar fraction D by HPLC, using hexane:ethyl acetate (72:28), yielded  $3\beta$ , $7\alpha$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -ergost-8-en-11-one (2) (6 mg) and  $3\beta$ , $7\beta$ -dihydroxy- $4\alpha$ , $14\alpha$ -dimethyl- $5\alpha$ -ergost-8-en-11-one (3) (1 mg). The spectral data of these novel compounds are given below.

 $3\beta_{\gamma}.7\alpha$ -Dihydroxy- $4\alpha_{\gamma}.14\alpha$ -dimethyl- $5\alpha$ -cholest-8-en-11-one (2) White needles; m.p.: 158–160°C;  $[\alpha]_{D}^{25}$  + 99.0° (c 0.33, CHCl<sub>3</sub>); UV (EtOH)  $\lambda_{max}$  256.0 ( $\varepsilon$  5900); IR (film)  $\nu_{\rm max}$  3374, 1642 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  4.35 (1H, dd, J=4.8, 1.1 Hz, H-7 $\beta$ ), 3.17 (1H, ddd, J=11.2, 9.6, 4.8 Hz, H-3 $\alpha$ ), 2.82 (1H, ddd, J=13.2, 3.6, 3.6 Hz, H-1 $\beta$ ), 2.64 (1H, d, J = 17.2 Hz, H-12 $\alpha$ ), 2.43 (1H, d, J = 17.2 Hz, H-12 $\beta$ ), 2.02 (1H, m, H-16), 1.86 (1H, m, H-15), 1.85 (1H, m, H-2), 1.80 (1H, m, H-6), 1.72 (1H, m, H-17 $\alpha$ ), 1.66 (1H, m, H-15'), 1.55 (1H, m, H-2'), 1.50 (1H, m, H-6'), 1.48 (1H, m, H-25), 1.41 (1H, m, H-4), 1.38 (1H, m, H-16'), 1.34 (1H, m, H-23), 1.35 (1H, m, H-20), 1.26 (3H, s, H-32), 1.2–1.1 (5H, H-1a, H-5a, H-23', H-24, H-24'), 1.03 (3H, d, J = 6.7 Hz, H-30), 1.02 (3H, s, H-19), 0.86 (6H, d, J = 6.7 Hz, H-21, H-26), 0.85 (3H, d, J = 6.7 Hz, H-27), 0.79 (3H, s, H-18); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) § 200.9 (s, C-11), 160.5 (s, C-8), 140.6 (s, C-9), 76.3 (d, C-3), 67.5 (d, C-7), 51.8 (t. C-12), 50.8 (s. C-14), 50.2 (d. C-17), 47.6 (s. C-13), 42.9 (d. C-5), 39.4 (t, C-24), 37.7 (s, C-10), 37.6 (d, C-4), 36.3 (d, C-20), 36.2 (t, C-22), 33.5 (t, C-1), 31.1 (t, C-2), 30.08 (t, C-15), 30.06 (t, C-6), 28.0 (d, C-25), 27.2 (t, C-16), 27.6 (q, C-32), 24.0 (t, C-23), 22.8 (q, C-26), 22.5 (q, C-27), 18.5 (q, C-21), 16.8 (C-18), 16.1 (q, C-19), 15.2 (q, C-30); HMBC cross-peaks C-1/H-5α, H<sub>3</sub>-19; C-3/H-2, H-2', H<sub>3</sub>-30; C-5/H<sub>3</sub>-19, H-7*β*; C-7/H-6, H-6'; C-8/H<sub>3</sub>-32, H-6, H-6'; C-9/H<sub>3</sub>-19, H-7*β*; C-11/H-12, H-12'; C-12/ H<sub>3</sub>-18; C-17/H<sub>3</sub>-18, H<sub>3</sub>-21; C-30/H-3 $\alpha$ ; EIMS m/z (70 eV) 444 (100) [M<sup>+</sup>], 429 (50)  $[M^{+}-15]$ , 420 (22)  $[M^{+}-18]$ , 401 (15)  $[M^{+}-43]$ , 292 (20), 263 (40), 237 (95); HREIMS m/z 444.3602 (calcd. for C<sub>29</sub>H<sub>48</sub>O<sub>3</sub>, 444.3603).

 $3\beta$ , $7\beta$ -Dihydroxy-4 $\alpha$ , $14\alpha$ -dimethyl-5 $\alpha$ -ergost-8-en-11-one (3) Amorphous solid:  $[\alpha]_{D}^{25} + 162.0^{\circ}$  (c 0.1, CHCl<sub>3</sub>); UV (EtOH)  $\lambda_{max}$  260.0( $\varepsilon$  7600); IR (film)  $\nu_{max}$ 3374, 1642 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  4.41 (1H, dd, J = 8.9, 8.9 Hz, H-7 $\alpha$ ), 3.07 (1H, ddd, J = 11.0, 9.8, 4.6 Hz, H-3 $\alpha$ ), 2.74 (1H, ddd, J = 13.2, 3.6, 3.6 Hz, H-1 $\beta$ ), 2.69 (1H, d, J = 17.2 Hz, H-12 $\alpha$ ), 2.54 (1H, d, J = 17.2 Hz, H-12 $\beta$ ), 1.23 (3H, s, H-32), 1.18 (3H, s, H-19), 1.03 (3H, d, J = 6.7 Hz, H-30), 0.9 (3H, s, H-18), 0.89 (3H, d, J=6.7 Hz, H-21), 0.88 (3H, d, J=6.7 Hz, H-26), 0.87 (3H, d, J=6.7 Hz, H-27);  ${}^{13}$ C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  200.8 (s, C-11), 160.0 (s, C-8), 141.9 (s, C-9), 76.0 (d, C-3), 68.7 (d, C-7), 52.0 (t, C-12), 49.9 (s, C-14), 50.8 (d, C-17), 47.6 (s, C-13), 42.9 (d, C-5), 39.4 (t, C-24), 38.1 (s, C-10), 37.1 (d, C-4), 36.3 (d, C-20), 36.2 (t, C-22), 33.8 (t, C-1), 31.7 (t, C-2), 30.05 (t, C-15), 30.06 (t, C-6), 28.0 (d, C-25), 27.3 (t, C-16), 27.3 (q, C-32), 24.0 (t, C-23), 22.8 (q, C-26), 22.5 (q, C-27), 18.6 (q, C-21), 17.8 (C-19), 16.9 (q, C-18), 15.1 (q, C-30); EIMS m/z (70 eV) 444 (91) [M<sup>+</sup>], 429 (78) [M<sup>+</sup>-15], 426 (23)  $[M^+-18]$ , 401 (17)  $[M^+-43]$ , 292 (19), 263 (50), 237 (100); HREIMS m/z444.3599 (calcd. for  $C_{29}H_{48}O_3$ , 444.3603).

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## References

- [1] J. Bellakhdar (1997). La Pharmacopée Marocaine Traditionnelle. Ibis Press, Maroc.
- [1] J. Benharref and J-P. Lavergne (1985). Bull. Soc. Chim. Fr., 5, 852–972.
- [3] S. Jabbouri, P. Chosson, P. Tisnes, R. Rao, P. Servin and J-C. Promé (1991). J. Chem. Soc. Perkin Trans., 1, 1935–1940.
- [4] R. Tanaka, K. Kasubuchi, S. Kita, H. Tokuda, H. Nishino and S. Matsunaga (2000). J. Nat. Prod., 63, 99-103.
- [5] T.-S. Wu, L.-S. Shi and S.-C. Kuo (2001). J. Nat. Prod., 64, 1121-1122.