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Y-Chromosome STR Haplotypes in Central-West African Immigrant in Spain Population Sample

KEYWORDS: forensic science, Y-chromosome, short tandem repeat, haplotype, forensic genetics, population genetics

Blood samples were obtained from 103 healthy unrelated black West African immigrant population in Spain from different countries situated to the south of the Sahara desert. The DNA was extracted using Chelex 100 protocol as described by Walsh et al. (1). The quantity of recovered DNA was determined using QuantiBlot® Human DNA Quantitation Kit (Perkin Elmer). DYS19, DYS389I, DYS389II, DYS390, and DYS393 were amplified as described by Gusmão et al. (2). The DYS385 amplification conditions complied with the methodology described by Schneider et al. (3). The PCR was performed in a Perkin Elmer 9600 thermocycler.

Allele nomenclature was proposed by Kayser et al. (4) and de Knijff (5) with the exception of the DYS389 locus. The nomenclature of this locus was according Gusmão et al. (2).

Electrophoresis was carried out on 4% polyacrylamide denaturing sequencing gels in a 377 automated system (Applied Biosystems Division/Perkin Elmer). Genotype classification was done using Genescan PCR analysis software with Local Southern Method and by side-to-side comparison with allelic ladders. These ladders were kindly provided by Dr. Carracedo (Institute of Legal Medicine of Santiago de Compostela), Dr. P.M. Schneider (Institut für Rechtsmedizin, Mainz, Germany). Haplotype diversity was calculated according to Nei (6). Analysis of Molecular Variance (AMOVA) was performed by Markov test using the Arlequin software 1.1 (7). Genetic distance matrix between populations were obtained by using the pairwise difference genetic distance.

A total of 98 different haplotypes were observed (Table 1), 93 of them being unique. The most common haplotype (DYS19: 15, DYS385: 16/16, DYS389I: 10, DYS389II: 28, DYS390: 21, and DYS393: 13) had frequency of 1.94% (h16, h21, h22, h38, and h54). Haplotype diversity for each of the markers is shown in Table 1. Haplotype diversity for all seven Y-specific STR loci in Central-West African Immigrant in Spain population was calculated to be 0.9990 (standard error: 0.0015).

Pairwise haplotype analysis using the population studied and other Iberian andAfrican populations (8,9) shows the valencian ($p = 0.000$) population distant from the rest of the populations.

The complete data are available to any interested researcher upon request.

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TABLE 1—*Y STR haplotypes in a black Central-West African immigrant population in Spain.*

Haplotype	n	F	DYS19	DYS389I	DYS389II	DYS390	DYS393	DYS385
h1	1	0,0097	15	9	26	22	13	14,15
h2	1	0,0097	15	10	28	21	10	17,19
h3	1	0,0097	14	11	27	21	14	16,16
h4	1	0,0097	15	10	28	21	13	17,17
h5	1	0,0097	16	10	28	22	13	16,27
h6	1	0,0097	14	10	27	21	13	14,17
h7	1	0,0097	13	11	28	23	12	15,16
h8	1	0,0097	16	10	28	22	13	16,17
h9	1	0,0097	14	10	28	21	13	16,19
h10	1	0,0097	15	10	28	21	13	16,18
h11	1	0,0097	15	10	27	21	14	18,18
h12	1	0,0097	16	8	25	21	13	14,14
h13	1	0,0097	15	10	27	21	13	17,17
h14	1	0,0097	15	10	28	21	14	15,15
h15	1	0,0097	16	10	29	21	14	14,16
h16	2	0,0194	15	10	28	21	14	14,16
h17	1	0,0097	16	9	26	21	14	16,18
h18	1	0,0097	14	9	26	22	14	16,20
h19	1	0,0097	15	10	28	21	13	14,18
h20	1	0,0097	16	10	28	21	14	13,16
h21	2	0,0194	15	10	27	21	14	16,17
h22	2	0,0194	16	10	27	21	14	15,16
h23	1	0,0097	16	10	28	21	13	17,17
h24	1	0,0097	15	10	26	25	13	13,16
h25	1	0,0097	15	10	27	21	13	17,19
h26	1	0,0097	15	10	27	21	13	13,15
h27	1	0,0097	15	10	29	21	13	16,17
h28	1	0,0097	15	10	27	20	13	14,16
h29	1	0,0097	15	10	27	21	14	14,16
h30	1	0,0097	15	10	27	21	14	16,16
h31	1	0,0097	15	8	26	21	13	16,16
h32	1	0,0097	15	10	26	24	13	13,15
h33	1	0,0097	15	10	27	21	14	15,20
h34	1	0,0097	17	9	26	23	13	14,14
h35	1	0,0097	17	9	27	21	15	17,17
h36	1	0,0097	15	11	28	21	15	18,21
h37	1	0,0097	16	10	27	21	13	15,17
h38	2	0,0194	15	10	28	21	13	16,17
h39	1	0,0097	16	11	29	21	14	15,16
h40	1	0,0097	17	10	28	22	15	15,16
h41	1	0,0097	16	10	28	21	14	17,18
h42	1	0,0097	15	9	26	21	14	16,16
h43	1	0,0097	15	9	26	21	16	17,18
h44	1	0,0097	15	10	29	21	13	15,16
h45	1	0,0097	15	9	26	22	13	15,16
h46	1	0,0097	16	10	27	21	15	16,18
h47	1	0,0097	15	10	27	22	14	17,18
h48	1	0,0097	14	9	25	25	13	19,19
h49	1	0,0097	16	10	28	21	13	17,18
h50	1	0,0097	15	9	27	21	13	15,16
h51	1	0,0097	16	10	28	21	14	16,16
h52	1	0,0097	15	18	28	23	15	16,16
h53	1	0,0097	15	10	29	21	13	15,18
h54	2	0,0194	17	10	27	21	14	16,16
h55	1	0,0097	15	10	28	21	14	15,16
h56	1	0,0097	15	10	28	21	13	17,18
h57	1	0,0097	16	10	27	20	14	15,15
h58	1	0,0097	15	11	28	21	14	15,17
h59	1	0,0097	16	10	29	21	14	16,16
h60	1	0,0097	16	10	28	21	13	14,15
h61	1	0,0097	15	9	27	22	13	13,15
h62	1	0,0097	15	8	25	21	13	16,16
h63	1	0,0097	15	10	27	21	14	16,18
h64	1	0,0097	16	10	29	21	14	16,17
h65	1	0,0097	13	10	28	24	12	15,16
h66	1	0,0097	15	11	27	21	14	18,18
h67	1	0,0097	15	10	27	21	13	15,18
h68	1	0,0097	15	11	28	21	14	17,17
h69	1	0,0097	14	9	25	22	14	16,18

continues

TABLE 1—*Continued.*

Haplotype	n	F	DYS19	DYS389I	DYS389II	DYS390	DYS393	DYS385
h70	1	0,0097	15	9	27	21	14	14,15
h71	1	0,0097	16	9	27	22	12	14,14
h72	1	0,0097	13	10	28	24	14	16.17
h73	1	0,0097	14	10	27	23	14	15,17
h74	1	0,0097	15	10	27	22	13	27,19
h75	1	0,0097	15	10	28	21	13	15,17
h76	1	0,0097	15	10	29	21	14	15,16
h77	1	0,0097	15	10	29	21	14	14,15
h78	1	0,0097	15	11	27	21	14	16,16
h79	1	0,0097	15	11	28	21	14	16,16
h80	1	0,0097	15	11	28	21	14	15,16
h81	1	0,0097	15	11	29	23	13	13,15
h82	1	0,0097	15	12	29	22	14	16,18
h83	1	0,0097	15	12	29	22	13	12,15
h84	1	0,0097	16	9	27	22	13	13,16
h85	1	0,0097	16	10	27	21	13	16,17
h86	1	0,0097	16	10	27	21	15	14,18
h87	1	0,0097	16	11	27	21	14	16,16
h88	1	0,0097	15	9	26	22	13	17,17
h89	1	0,0097	16	11	27	22	13	16,17
h90	1	0,0097	16	11	28	21	15	17,17
h91	1	0,0097	17	9	26	22	13	17,19
h92	1	0,0097	17	10	27	20	14	15,16
h93	1	0,0097	17	10	28	20	15	16,16
h94	1	0,0097	17	11	28	22	13	16,16
h95	1	0,0097	17	11	28	22	13	15,16
h96	1	0,0097	17	11	28	22	13	15,18
h97	1	0,0097	17	11	28	22	13	16,15
h98	1	0,0097	17	1	28	22	13	17,17
HVD			0,6463	0,5888	0,7132	0,5313	0,6164	0,9359

* N: number of haplotype; n: individuals observed for each haplotype; HVD: haplotype diversity value; F: frequency for each haplotype in 103 individuals.

Correspondence

Commentary on: McBride DG, Dietz MJ, Vennemeyer MT, Meadors SA, Benfer RA, Furbee NL. Bootstrap methods for sex determination from the os coxae using the ID3 algorithm. J Forensic Sci 2001;46:424–428.

Sir:

In a recent article in this journal, McBride et al. (1) present a ground breaking forensic anthropological analysis based on an early "machine learning" algorithm. While there is much to commend in this article, as a first application there are also the inevitable points that raise questions relevant to future work in forensic anthropology. In this letter we raise a few of these issues.

Typically there are two goals to forensic anthropology. The first goal is that of estimation, in which a profile is built from the skeletal remains of an unidentified individual in the hopes that said remains may eventually be identified. As a statistical pursuit, such estimation needs to be probabilistically based. It is not enough, and indeed it is misleading, to state that the remains belonged to a "white male who was 42 years old and 6 feet tall." Continuous variables (such as age and stature) need to be stated as "highest posterior densities," while categorical variables (such as "race" and sex) should be given with their posterior probabilities. As the American public continues to be fed a diet of both reality-based and fictionalized accounts of detective and forensic anthropological cases, it is important that we provide information couched in the ambiguities of the science. Failure to do so may lead to missed opportunities at identification.

The second goal of forensic anthropology is to provide statistical evidence in the case of a putative (or so-called "positive") identification. This again is a probabilistic problem, as the anthropologist (minimally) needs to present a likelihood ratio (2). While there are many expert forensic anthropologists who are quite good at providing unbiased and efficient point estimates and categorical statements, if more "objective" forms of data (such as DNA) go challenged in the courts the day cannot be far off when an expert forensic anthropologist will face similar challenges on presenting evidence in an identification-based case. Forensic anthropologists who have had considerable training and much prior experience can, and often do, learn to glean a surprising amount of information from a single skeleton and its context. Machines (i.e., computers) are not particularly good in such situations, and it is doubtful that a jury will follow an argument based on "artifical intelligence" or presented on behalf of a "learned machine." Neither do we expect that judges will relish instructing a jury on how to interpret evidence presented on behalf of an "expert system," rather than by an expert.

All of this is not to say that computers have no place in the future of forensic anthropology. It is precisely in the domain of probabilistic statements that computers can be "taught to think" (i.e., programmed) quite effectively, while humans are understandably poor at processing the often massive amounts of data needed to calculate probabilities. As an early computer algorithm designed to solve discrete problems, the ID3 algorithm that McBride et al. used is not well suited to providing probabilistic statements. Newer statistical methods that bear some similarities to ID3, such as Bayesian CART (for "classification and regression trees") (3) can provide probabilistic statements. There are also now a large number of alternatives to ID3 (4), and older parametric models such as probit are also a possibility (5). And while we would not relish explaining Bayesian CART or cumulative probit models to a court, we would feel better about doing so than we would providing a "cut-and-dry decision" from ID3. A further problem with ID3, which is alluded to by McBride et al. ("ID3's prioritization of attributes may be viewed as problematic," p. 428) is that it does not tend to "think" in a manner similar to how expert forensic anthropologists analyze cases. ID3 is a so-called "greedy" algorithm, in that it partitions data sets by sequentially moving through attributes in order of information gain. In contrast, forensic anthropologists tend to use a "weight of information" approach, where they form opinions about cases based on agreement between a number of different attributes. In the following, we describe a recent large "experiment" in estimation of sex from the os coxae as a contrast to McBride et al.'s study.

In June of 1998 we scored 793 os coxae from the Terry Collection (the source for McBride et al.'s 115 cases) as part of a larger study on estimation of age-at-death from skeletal remains (NSF SBR-9727386, see http://konig.la.utk.edu/paleod.html). We describe our examination and scoring methods in detail here, as they are pertinent to how we made determinations of the sex of individuals. Because we collected pubic symphyseal ageing data that is scored differently for the sexes (6,7), it was necessary to "know" the sex of each individual when we were scoring. However, in an actual forensic context sex typically would not be known, and so we estimated the sex for each of the 793 cases using the three Phenice (8) characteristics that also appear in McBride et al.'s analysis. We scored each characteristic as "F," "F?," "?," "M?," "M," or unobservable. Now there is a bit of fiction involved in saying that we used the Phenice characteristics (and *only* those characteristics) in making determinations of sex. All os coxae were scored blind and independent of any other bones from the skeleton, but like in McBride et al.'s study the observer had access to the entire os coxae, and so may have used other (unrecorded) criteria. That said, we should also point out that our study was done principally over eight days, with the least number of os coxae scored on any one of these days being 49, and the most being 152. In addition to scoring the Phenice characteristics we were collecting pubic symphyseal and auricular phase data, as well as age "indicator" data from the cranium and long bones. We consequently spent considerably less time with each skeleton than would be true in a forensic context, and so it is unlikely that we used anything more than the Phenice characteristics with any regularity. In order to expedite the study we divided tasks so that the second author, who had the most experience in scoring auricular surfaces, scored most of the os coxae. Consequently, he was responsible for sexing about 90% of the bones, while the other authors each did about 5% (for those wanting more detail, the complete data set is available by anonymous ftp from the web-site listed above).

Instead of using ID3 with the Phenice characteristics we used a more modern program, Polytomous Logistic regression trees with Unbiased Split (PLUS) (9) to form a decision tree from the three

Phenice characteristics. PLUS is currently available from http://recursive-partitioning.com/plus/. We treated the characters as numeric, with "F" = 1, "F?" = 2, "?" = 3, "M?" = 4, and "M" = 5. Unobservable traits were handled using "nodewise imputation" and for cross-validation we used individual cases (i.e., a "leave-one-out" strategy). The program found that the lowest cross-validated error rate occurred when there was a simple split on subpubic concavity, with "F" and "F?" going to the "left" and "?," "M?," and "M" going to the "right." This simple tree gave a cross-validated error rate of 2.4%, misclassifying 8 of the 361 actual females in the sample as males and 11 of the 432 actual males as females. This misclassification rate is lower than all of the "mean error percents" given in McBride et al., most of which were found using a large number of attributes, as versus the single attribute subpubic concavity. While again, it could be argued that our scoring of subpubic concavity was influenced by observing other unrecorded attributes in the os coxae (and was not made independent of the other two Phenice characteristics), such an argument cannot be made for a study by Sutherland and Suchey (10). In a study of 1284 pubic bones that had been removed at autopsy, and in which only Phenice's "ventral arc" could be scored for its presence/absence, the authors misclassification rate was 4%, again lower than all of McBride et al.'s mean rates.

Because decision trees from PLUS, like those from ID3, are univariate, we also consider the "Linear Machine Decision Tree" (LMDT) algorithm. LMDT (11) uses a multivariate model (discriminant analysis) in a decision tree setting, and allows for "pruning" of the tree so that it does not become overly complex ("bushy"). The source code for LMDT is currently available from http://yake.ecn.purdue.edu/~brodley/software/lmdt.html. We applied LMDT, again using node-wise imputation for missing data. Ultimately, LMDT misclassified 9 of the 361 actual females as male (one more misclassification than PLUS) and 6 of the 432 actual males as female (5 less than PLUS with cross-validation, or 4 less than PLUS using the biased "plug-in" rule). For complete cases (i.e., cases with no missing data) LMDT first makes a split simultaneously on the basis of the ventral arc and sub-pubic concavity. Cases where the sub-pubic concavity is scored as "F" and the ventral arc as no more male than "M?," or the sub-pubic concavity is scored as "F?" and the ventral arc as no more male than "?," or the sub-pubic concavity is scored as "?" and the ventral arc as no more male than "F" are all classified as "female." The remaining cases are then split again using the sub-pubic concavity and ischio-pubic ridge. Cases that have both features scored as "M" are classified as male. Cases that do not have both features scored as "M" go through further splits, but in the interest of brevity we do not describe them here.

We can also compare our results using decision trees to what we obtained simply by "mentally processing" the information from the Phenice characteristics. Some points are in order here before we turn to this comparison. First, there were three clerical errors we found after the fact. We correctly classified two individuals by sex, even though we have no recorded Phenice characteristics from these individuals. As we have scores for their pubic symphyseal development it is likely that we simply failed to record the Phenice characteristics for these two individuals. There is an additional clerical error, in that one individual for whom we scored all three characteristics as unambiguously female we also have classified as a male. Outside of these three errors, there are four cases where our opinion on sex classification is not consonant with the Phenice characteristics, although in all four instances we correctly identified the sex. In one actual male, where we scored the ventral arc as "F," the subpubic concavity as "F?," and the ischio-pubic ridge as "M" we ultimately classified the individual as male. For two actual males where we scored the ventral arc as "M" and the other two characters as "F?" we identified the individuals as males. Finally, for one actual male where we scored the ventral arc as "F?," the subpubic concavity as "M?," and the ischio-pubic ridge as "?," we ultimately identified the individual as a male. In all four cases, we must presume either that we were drawing on other attributes, or that our visual weighting is not well reflected in the three Phenice scores. Aside from these exceptions, all of the sexes we assigned in June, 1998 were done by taking the majority characteristic. In other words, if two of the characteristics were "F" while one was "M" or "?" we classified the individual as a female, and we treated unobservable characteristics as uninformative. On this basis, we ultimately misclassified 7 of the 361 actual females in the sample as males (2 less than LMDT) and 5 of the 432 actual males as females (1 less than LMDT).

Now, how might we use the above-described study in forensic settings? Our "mental processing" did not use the sample itself to derive rules, so our observed misclassification rates should be applicable to any new samples. By Bayes Theorem, if we assume that an unidentified case is as likely to be from a male as from a female, then upon "determining" sex on the basis of the Phenice characteristics we should say that individuals we identify as female have a 0.9983 posterior probability of actually being female (equal to (354/361)/(354/361 + 5/432)). Similarly, those we call "male" have a 0.9808 posterior probability of being male. In an identification case, if the identification (external to the osteological evidence) is for a female, and we suggest on the basis of the Phenice characteristics that the individual has the morphology of a female, then the likelihood ratio (assuming an even sex ratio in the "population at large") is 1.9667, while for a male who has male Phenice characteristics the likelihood ratio is 1.9615. While these are very weak likelihood ratios (because there are only two sexes, which we have assumed are equally frequent), they could be combined with likelihood ratios from other osteological evidence in order to "sharpen" the posterior odds. That these likelihood ratios are nearly 2.0 is because the level of misclassification is extremely low. "Machine learning" methods could presumably be applied to other osteological classification problems, where the number of categories is greater than two. While such classifications would be more informative in identification cases, we also suspect that the level of misclassification could be unacceptably large. But for the present setting of sex identification from the os coxae, we do not find that "machine learning" methods offer much beyond what a reasonably well-trained and experienced osteologist can provide. In point of fact, McBride et al.'s ID3 based analysis did not perform nearly as well as we did, calling into question not only the algorithm, but also the quality of the attribute scores from which the program generated its decision tree.

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Authors' Response

Sir:

We are pleased to have generated interest in our recent paper (1). Konigsberg, et al. (2) present a valuable contribution to the use of machine learning algorithms in questions relevant to forensic science. Their work points out the increasing importance of presenting the conclusions of osteological analyses in probabilistic terms, but the direct criticisms of our work distort its central concept.

Our study (1) proceeded from the point of view that, in order to obtain an accurate and informative analysis of skeletal remains, one must first decide what, specifically, to analyze. Expert skeletal analysts have the benefit of extensive training and experience to guide and support their conclusions, and will have refined their own preferred techniques over many years. In contrast, skilled, but nonexpert, analysts often face difficulties when compelled to select among many methods and criteria, as may be the case with, for example, fragmentary or otherwise unusual material. Phenice (3), Sutherland and Suchey (4), Rogers and Saunders (5), and most recently, Konigsberg, et al. (2), show that small groups of attributes can be quite accurate. Small sets of attributes have an added advantage over large sets in being less likely to include unobservable attributes. However, no attribute set is necessarily optimal in every instance.

McBride, et al. (1) presented ID3 in a useful, nonarbitrary and repeatable procedure for identifying good subsets of attributes. To show that the chosen attributes worked as well as all attributes together, training sets of 70% of the sample were drawn randomly with replacement and tested against the remaining 30%. Relative error rates averaged over ten trials were cited for each attribute suite as proof of success in this regard. We stated that "The selected attribute suite of preauricular sulcus, sciatic notch, and subpubic concavity should provide good results when scored as indicated in Table 1," but did not state a specific level of accuracy that one should expect in a different context. Because we developed the attribute suites with a bootstrapped training set/test set protocol and presented ten-trial averages for each attribute set, we regard our results as robust.

The 31 attributes used in the study were based on the techniques of three widely recognized experts in skeletal analysis (see Table 1 (1)). They are therefore partly redundant in that some attributes require different criteria for scoring the same os coxae features. ID3 demonstrated sensitivity to semantic differences in the definitions of each attribute. Out of approximately 90 trials, some "versions" of attributes were never preferentially chosen by ID3, and others commonly were. Similarly, several attribute descriptions commonly appeared as second or third branches on decision trees; never first, never last, suggesting those attributes contained information useful in segregating cases that remained ambiguous after the first split. The sensitivity of ID3 to these subtleties may reflect intraobserver variability in applying each criterion, or it may suggest a broader interpretation, for example, that Expert C's "subpubic concavity" criteria capture more information than Expert B's. If taken in context, it does not necessarily, as Konigsberg, et al. state, "call into question not only the algorithm, but also the quality of the attribute scores from which the program generated its decision tree." (2). Instead, it suggests that ID3 may be useful in examining relative effectiveness of individual attributes, questions of inter- and intraobserver variability, and the semantics of attribute definitions.

To address the question of accuracy with results more directly comparable to Konigsberg, et al. (2) than our original study presented, we obtained the larger Terry Collection data set made available on the Internet by Konigsberg, as well as the LMDT (6) and PLUS (7) software (see Konigsberg, et al. (2) for URLs). We also ran new trials using ID3 with our original, raw sample of 115 Terry Collection individuals. Using a "leave-one-out" jackknife procedure with our sample of 115 (35F, 80M), we tested our preferred attribute suite of preauricular sulcus, sciatic notch, and subpubic concavity (1), as well as attribute sets we designated to represent those recommended by Phenice (1,3). We also intended to test our 115 sample and one training set/test set trial under PLUS and LMDT. However, LMDT has not run successfully on our system, and technical support is no longer offered by its author (Carla Brodley, e-mail communication, 2001); results are given only for PLUS. Finally, using ID3 we tested 600 individuals (total 300F, 300M) from the Konigsberg Terry Collection sample. It was necessary to divide the sample of 600 into three trials of 200 each (100F, 100M) because our implementation of ID3 is an MS-DOS program that can only access enough memory for slightly more than 200 cases at once. We must stress that this is a limitation of our ID3 software, not of the algorithm itself.

Using the Terry Collection data provided by Konigsberg (Phenice's attribute set) and a leave-one-out protocol, ID3 misclassified a total of 8/300 females (2.6%) and 7/300 males (2.3%), for an overall cross-validated error rate of 15/600, or 2.5%. In one trial using our 115 sample with ID3, our preferred suite of attributes misclassified 3/35 females and 1/80 males, for a total error rate of 4/115 (3.4%). The "Phenice" attributes designated in our original paper (a "generic" set, comprised of one attribute definition from each expert; see Table 1 (1)) misclassified 8/35 females and 4/80 males. For comparison with our generic "Phenice" suite, we re-ran the trial three times, each time using the three "Phenice" attributes as defined by each expert. Interestingly, misclassification rates were slightly different for the various definitions of the "Phenice" attributes: Expert A's suite misclassified 8/35 females and 2/80 males, Expert B's suite misclassified 8/35 females and 4/80 males, and Expert C's suite misclassified 5/35 females and 1/80 males. Each trial with the "Phenice" attributes generated a distinct, albeit similar, tree in which cases were ordered differently and some different cases were misclassified, despite having been developed from the same data. This further suggests that ID3 is sensitive to subtle, qualitative differences in scoring, as noted above.

We tested PLUS using our 115 sample and one arbitrarily chosen training set/test set sample that was originally drawn for the

ID3 trials. With 115 cases, our preferred suite of attributes misclassified 1/35 (2.8%) females and 4/80 (5%) males. The "Phenice" attributes misclassified as follows: the generic attributes misclassified 3F and 9M; the Expert A attributes misclassified 10F and 2M; the Expert B attributes misclassified 12F and 3M; the Expert C attributes misclassified 5F and 2M. In the training set/test set trial with PLUS, our preferred set of attributes misclassified one female and two males, with a cross-validated error rate of approximately 2.5%. The generic "Phenice" attributes misclassified eight females and one male, with a cross-validated error rate of 11.2%.

The brief comparisons presented above do not suggest that ID3 is inferior to PLUS. Although our data sets returned slightly weaker results in some trials, it should be noted that the ossa coxarum were originally scored by one individual with the specific objective of duplicating the recommendations of three different experts in skeletal analysis. That ID3 and PLUS both detected subtle differences among attribute suites (reflected in varying accuracy rates for the "Phenice" trials) suggests the scorer's original objective was met. In regard to our "poor" results with ID3 and the Phenice characteristics, we are not surprised that Konigsberg, et al. (2) obtained a better result using a sample nearly seven times larger. Methods do better on the samples from which they are developed. Phenice (3) obtained his results using 275 individuals from the Terry Collection. Our results (1) were obtained from a Terry Collection sample of 115 individuals and validated with a rigorous training set/test set protocol averaged over 10 trials. Since it is quite likely that substantial portions of our and Phenice's samples are contained within the Konigsberg Terry Collection sample, we would be concerned if Konigsberg, et al. (2) had not obtained a better result. Under the more directly comparable protocols presented here, the misclassification rates of 2.6% F and 2.3% M for ID3 using the Konigsberg Terry Collection sample evaluate favorably with the errors of 2.2% F and 2.5% M (PLUS), and 2.4% F, 1.3% M (LMDT) reported by Konigsberg, et al. (2).

We agree with Konigsberg, et al. (2), that there is a need to develop probabilistic statements for categorical variable estimations, such as sex. Either ID3 or PLUS should be useful in these and other applications of machine learning algorithms to skeletal analyses. Several other algorithms and/or software based upon them were noted by Konigsberg et al. (2) and McBride, et al. (1). They are also readily available on the Internet and merit evaluation in future studies. It is important to note in closing that none of these programs constitute expert systems in themselves. They are tools intended to assist the development of rule sets, or decision trees, such as those discussed above, which are the building materials of expert systems. A finished expert system may incorporate several hundred rule sets, much refined after consultation and validation with several human experts. Such a hypothetical system would produce skeletal analyses equivalent to analyses done by the experts who contributed to its development.

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Commentary on: Dou C, Bournique J, Zinda M, Gnezda M, Nally A, Salamone S. Comparison of Rates of Hydrolysis of Lorazepam-Glucuronide, Oxazepam-Glucuronide and Temazepam-Glucuronide Catalyzed by E. Coli β-Glucuronidase Using the Online Benzodiazepine Screening Immunoassay on the Roche/Hitachi 917 Analyzer. J Forensic Sci 2001;46(2):335–340.

Sir:

In their article, Dou, et al. report catalytic rates and Km values for the hydrolysis of lorazepam-glucuronide, oxazepam-glucuronide, and temazepam-glucuronide by *E. Coli* β-glucuronidase. The authors report that the purity of the glucuronide reference materials, purchased from Alltech, is greater than 90%; however, they do not mention if they actually validated the concentrations of these reference materials prior to use or merely reported the manufacturer's labeled concentration. Although Alltech lists the concentration of these reference materials as 1 mg/mL in their catalog, they do note that due to limited supplies of pure standard, these solutions are not quantitative.

In a previous study (2) we reported on the validation of (R,S) lorazepam- and oxazepam β-glucuronide primary reference materials for hydrolysis and quality assurance controls. Using HPLC analysis and GC/MS analysis following both acid and β-glucuronidase hydrolysis, we found that the lorazepam β-glucuronide material was >95% pure, but the oxazepam β-glucuronide material was only 54% pure. At the time the manufacturer stated that their reference materials were highly purified (>95% pure) but were intended for use only as qualitative standards. The materials were semi-quantitative and were not intended for use as quantitative standards. In order to perform the cross reactivity and kinetic studies described by Dou, et al., the concentration of the substrates must be known in order to obtain valid results. We are concerned that the authors made no mention of the validation of the concentrations of these glucuronide reference materials prior to performing these studies. Without the validation information for the reference material purity data, it is unclear whether the differences in the hydrolysis rates and Km values are a result of structural differences or an error in the substrate concentration.

Although the benzodiazepines are stereospecifically conjugated so that only one isomer (the β-glucuronide) exists in biological systems, both oxazepam and lorazepam are racemates since the 3 carbon on the diazepine ring is chiral. Therefore, both the R and S isomers of the β-glucuronide are present in urine, plasma as well as these reference materials. Depending on the source of the β-glucuronidase, the S and R isomers can be hydrolyzed at different rates (3). It has been reported that the β-glucuronidase from *E. Coli* is very selective hydrolyzing the S isomer 446 times faster than the R isomer (at 37°C). Therefore, the relative concentrations of the S and R isomers may impact the initial hydrolysis rates of these reference materials. We reported that relative concentrations of the S and R isomers of oxazepam β-glucuronide were 61.0% and 39.0%, respectively and 54.5% and 45.5% for lo-

razepam β-glucuronide (2). It is not known if the relative concentrations of the isomers in the reference materials have changed over time.

As stated in our previous study, Alltech's glucuronide materials can be valuable as hydrolysis controls in method development and routine benzodiazepine analyses, but each laboratory must validate the purity of these glucuronide reference materials prior to use.

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Authors' Response

Sir:

The authors O'Neal and Poklis bring up valuable points that are thoroughly discussed in their paper (*Forensic Science International* 79, (1996) 69–81). They question whether or not validation of the glucuronide reference materials was performed prior to the experiments conducted to determine the catalytic rate constants for the hydrolysis of lorazepam-glucuronide, temazepam-glucuronide, and oxazepam-glucuronide. We wish to acknowledge the fact that although no in-house analytical determination for purity of the glucuronides was performed, we did obtain the analytical Standard Certificates of Quality from Synthetic Technology Corp (manufacturer of the material). The report by O'Neal and Poklis in 1996 induced Synthetic Technology Corp to reexamine their methods of preparation and characterization of the benzodiazepine glucuronide material.

The reported purity was >90%, >95% and >95% for temazepam-glucuronide, lorazepam-glucuronide, and oxazepam-glucuronide, respectively. This was based on HPLC analysis. In addition (precipitated by the concerns of O'Neal and Poklis) we contacted the vendor and obtained the following information[1]:

- The temazepam-glucuronide used in the studies was initially bottled in August 1993 from stock synthesized in April of that year. It was the vendor's first synthetic batch of this material. The vendor used 7.0 mg of exhaustively purified (by NMR) stock and diluted by weight into HPLC grade methanol to a resulting concentration of 0.1mg/mL. The chromatogram supplied with the material was checked and found to be accurate, indicating the primary "contaminants" being the minor "satellite" sugar enantiomers of the R/S mix. The stated purity for this lot of material was given as greater than 90%.
- The Lorazepam Glucuronide used in the studies was prepared in November 1995 as a part of a lot of 108 ampoules. It was prepared in a similar manner as described above with 10.9 mgs of exhaustively purified stock diluted in HPLC grade methanol and immediately sealed in silylated vials. Both the purity and the calculations of concentration have been verified to be correct.
- The oxazepam glucuronide used in the kinetic studies was prepared in December 1997 as a part of 108 ampoules. 11.74 mgs of exhaustively purified oxazepam glucuronide stock was diluted in HPLC grade methanol and immediately sealed in silylated vials. Again, both the purity and the calculations of concentration were reviewed and determined to be accurate. This material is probably the purest of all three.

Of the three lots the Temazepam is the oldest material, of the lowest (>90%) original purity at the time of shipment. The "primary" impurity was determined to be the satellite mix of enantiomers based upon the four possible combinations of the coupling of the sugar with the hydroxylated benzodiazepine.

The biggest concern we had regarding the purity of the material was with the stability of the glucuronidated conjugate. Upon aging and degradation the free aglycone, or similar chemical byproducts would be expected to be present. We tested this purity by antibody cross-reactivity. If there was a substantial amount of non-glucuronidated material, the immunoassay (which has low selectivity to the glucuronides and high cross-reactivity to the free drug) would have picked it up. Our study indicated that the lots did not contain significant amounts of this expected contaminant.

The main focus of the study was to compare hydrolytic rates of several benzodiazepine-glucuronide conjugates under various conditions. If the material was not pure the catalytic turnover (Kcat) and effects of temperature, matrix and pH on Kcat would not significantly change the values since the substrate is used in excess and only product formation is monitored. The Km would, however, be effected and conclusions concerning the binding affinity of each glucuronide to the enzyme would be in question.

O'Neal and Poklis also commented on the fact that depending on the source of the β-glucuronidase, the rates of hydrolysis of the R and S isomers vary. They state that β-glucuronidase from *E. coli* hydrolyzes the S isomer 446 times faster than the R isomer (at 37°C). In the case of our studies we were measuring initial rates (two minute reaction) so that the selectivity for one isomer over the other would not be observed. The enzyme may have been more selective for the S isomer and if so the resulting product would have been the corresponding enantiomer. In the light of this selectivity it would have been better to use the pure diastereoisomers for this study.

The authors raise good points about the chemical and stereochemical purity of the glucuronide conjugates. While we feel confident of the purity provided by the information from the manufacturer and by the testing that we did in house, it still would have been prudent (in light of the O'Neal and Poklis paper) to additionally test for the purity using another physical method.

Reference

1. Benzodiazepine glucuronide standards information was obtained from Dr. Mark Hagadone (Synthetic Technology Corp.) via personal communication.

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Commentary on: Lee GSH, Brinch KM, Kannangara K, Dawson M, Wilson MA. A methodology based on NMR spectroscopy for the forensic analysis of condoms. J Forensic Sci 2001;46(4): 808–21.

Sir:

I read with great interest the paper by Dr. Garry S. Lee et al., "A Methodology Based on NMR Spectroscopy for the Forensic Analysis of Condoms." I believe their NMR method may have potential applications far beyond what even the authors envision. One example is when questions are raised regarding the veracity of DNA evidence. Some recent reports (1–3) have explored the confusion that can arise in DNA evidence when the suspect has attempted to outwit it by selling, exchanging, or mixing semen samples.

With NMR we may now have a simple method (yes, the instrumentation is expensive, but the extraction and method are simple) that in one test can provide a profile of residues one might obtain from different brands of condoms. In one case (1) where there was a seminal fluid stain on the victim's blue jeans, whose DNA did not match that of the suspect, an unused portion of the stain could have been cut-out and extracted with hexane. Some other areas (where no seminal fluid or any other type of stain was visible) of the blue jeans could have been used as controls and extracted separately with hexane. After the hexane had been evaporated off, the residues could have been dissolved in an appropriate NMR solvent and examined. Comparison of the peaks from the controls and the seminal fluid stain would tell us which peaks were due to substances generally present on the jeans (for example, detergent residues) and those that might have been associated with condom traces. It is very unlikely that seminal fluid would interfere with this comparison. It does not interfere with identification by FTIR and/or desorption chemical ionization mass spectrometry (4) of the silicone oil, polydimethylsiloxane (PDMS), after extraction with dichloromethane and hexane is an even less polar solvent.

By comparison with a library of the NMR profiles of various condom brands (the authors correctly point out that such a library would have to be maintained), it might not only be possible to identify various components (PDMS, polyethylene glycol, nonoxynol-9, etc.), but also be feasible to at least profile a condom from a particular manufacturer (even if there were several different brands made by this manufacturer.)

I believe strongly in the potential of this NMR method; however, I do feel it is necessary to point out a small omission in the paper. In the first paragraph at the top of page 809 the authors state: "There are two types of lubricants used on condoms—those based on PDMS and those using polyethylene glycol 400 (PEG)." Actually I am aware of at least two others. Perhaps most Trojan brands (Carter-Wallace) are not available in Australia (the authors only list the "Naturalamb" Trojan brand.) I do not know their exact market share, but various Trojan brands are large sellers in the USA. Although Carter-Wallace uses PDMS in those brands that are advertised as just "lubricated", their chemists feel that PDMS and nonoxynol-9 are incompatible. Therefore in those brands that contain this spermicide, a water-soluble gel-type lubricant is used. This lubricant contains many ingredients, but by far its major one is propylene glycol (please note that this is not the polymer PEG). In addition, I believe there are several brands sold outside the USA that contain glycerol as a lubricant.

This minor correction in no way detracts from the excellence of the paper. In fact, the greater diversity among condom lubricants can only increase the discriminatory value of their NMR method. The many components in the gel-type lubricant used by Carter-Wallace should especially provide a unique NMR signature. Propylene glycol and glycerol are both quite polar and very water-soluble; the authors might wish to try extracting these condoms with a polar solvent and dissolving the residue in a polar NMR solvent.

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Authors' Response

Sir,

In a previous paper (1), we examined the technique of solid state and solution nuclear magnetic resonance (NMR) spectroscopy for the forensic analysis of Australian condoms. We presented a methodology that was capable of distinguishing condoms from different manufacturers based mainly on the comparison of the ^{1}H NMR spectrum (condom spectral fingerprint) of hexane extracted portions. This technique made use of the different compositions of chemical additives, preservatives, spermicides, and lubricants that were added to the condom by the manufacturers.

In his comment of our work, Blackledge (2) has made a number of points. He comments that condom trace evidence can provide clues into crimes in which the suspect has been cunning enough to try and outwit such as buying or exchanging semen by criminals to fool DNA analysis. In our paper we could have listed a myriad of other examples where DNA evidence may be a problem, involving bizarre sexual practices and multiple rapes. However, we wanted to stress the point that of the currently used techniques in the analysis of condom trace evidence, NMR is by far the most successful and useful in discriminating individual brands. It is well established that DNA analysis should not be considered the "be all and end all" of forensic science and our work assists here.

The second point is more implied and was mentioned in our original manuscript. However, it is still worthy of reiterating, i.e., that trace evidence of this kind can assist investigators by providing associative evidence and thus provide a link to the crime or it can aid in proving that a crime did actually occur.

The third point that there exists four types of lubricants and not two is trivial but points to the need to internationalize the work. Our research and personal communications with representatives of various companies revealed that only two lubricants, polydimethylsiloxane (PDMS) and polyethylene glycol 400 (PEG) based, were used at the time of the research in Australian condoms. The Trojan brand by Carter-Wallace, which is said to contain a propylene glycol based lubricant, is not available in Australia. In fact, Carter-Wallace only produces one brand in Australia, Bikini (3), which as afar as we can ascertain is a non-lubricated condom (3). Nevertheless, it was so rare that we were not able to obtain a sample at the time of the investigation. However, this does not mean that a condom, possibly Trojan, or one which contains a glycerol-based lubricant cannot make its way into Australian society. The ease of mail order buying and internet shopping coupled with the many visitors coming into the country at any one time provides plenty of opportunities for these condoms to enter the country. Moreover, propylene glycol and glycerol based lubricants are available in Australia but as additional or separate gels or creams that may be applied to the condom. We also understand that in the course of development, new lubricants and spermicides will be developed and thus have stressed in our original manuscript the need to keep the database updated (a point also agreed on by Blackledge).

To expand this point ad nauseam, in addition to polydimethylsiloxane, polyethylene glycol 400, glycerin and propylene glycol, we also know of attempts to use polypropylene glycol, polyisobutene, polyoxyethylene, behenic acid, behnyl alcohol, sorbitol (4) as well as mixtures of alcohols and acrylic polymers (5–7) as lubricating agents in condoms. Okamoto Industries (who manufacture the Liaison brand) and Sagami Rubber (Duo and Doms brands), both in Japan, have experimented with these new lubricants (5–7). In addition, organometallic zinc compounds are currently being tried as spermicides (4). For completeness, it should also be mentioned that manufacturers such as Sesicon Corporation have produced condoms using new polymer backbones that are yet to be available in Australia. Tactylon from Sesicon Corporation is made from a styrene-ethylene-butylene-styrene polymer, and there are others in the developmental stages that may become available in the near future. It goes without saying that all of these products, when released to the market, must also be included into the database.

Finally, Blackledge also cites an example of a smeared semen stain that may be extracted and tested for components of the condom lubricant. Although he uses this as an illustrative example, this may be construed by those unfamiliar to NMR, that it is a sufficiently powerful enough technique for the analysis of lubricant traces from condoms in semen stains. Unfortunately, currently this is not true. In such small dilute samples, the strong lubricant base may be detected, but the smaller trace components, which distinguish the individual condoms, will not. However, NMR is a versatile technique and many different experiments may be executed to obtain different kinds of information. Work is also currently underway to improve the detection limits for condom trace analysis by NMR, making use of the new pulse field gradient techniques to suppress the strong lubricant and/or solvent signals. Moreover, micro-NMR techniques are now becoming available. These techniques are capable of producing high-resolution spectra from nano-liter volume samples (8,9) giving hope to forensic scientists that analysis of this kind may be possible in the near future.

We reiterate that our paper is a somewhat introductory investigation and covered the popular and easily obtainable condoms in NSW, Australia. Information from the Therapeutic Goods Administration (TGA) details that 21 brands (79 different condoms) are available for retail sale in Australia (3). In our investigation, we have only looked at 38 condoms from 12 of the brands. Further research is currently underway to establish a complete international database with all available condoms, including novelty ones because although they are not Food and Drug Administration (FDA) approved, they can still be used to hide or remove bodily fluids. An international database such as this would be invaluable to forensic scientists. A pattern recognition computer system is also being developed for the database (10).

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