



Aplysiopsenes: an additional example of marine polyketides with a mixed acetate/propionate pathway

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ABSTRACT

First chemical study of the Hermaeidae sacoglossan *Aplysiopsis formosa* from Azores led to aplysiopsenes, α -pyrone polyketides with a mixed acetate/propionate structural pathway.

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Polypropionates displaying a pyrone ring are quite rare in nature. These are detected in some *Streptomyces* bacteria¹ and fungi of different genera.² Surprisingly, the biosynthesis experiments proved two different pathways: a mixed acetate/propionate in *Streptomyces*³ and methylation of a polyacetate chain in fungi.^{4,5} Related compounds were found in some marine organisms and, in particular, in sacoglossan molluscs.⁶ Origin and function of these molecules are challenging topics for international debates. Some authors⁷ suggested that these are produced by symbiotic microorganisms living in the molluscs, whereas others⁸ proved that γ -pyrones in sacoglossan polypropionates can catalyse the production of highly reactive singlet oxygen. Simple α - and γ -pyrone polyketides were found in four sacoglossan species: *Cyerce nigricans*,^{9,10} *Cyerce cristallina*,^{11–13} *Ercolania funerea*¹⁴ and *Placida dendritica*.^{15,16} All these molluscs, belonging to the superfamily Limapontioidea, are characterised by dorso-lateral appendages, named cerata, where the chloroplasts sequestered from the dietary algae can remain active for many months. The cerata can be easily autotomised in case of external stress including the escape from the predators.⁶

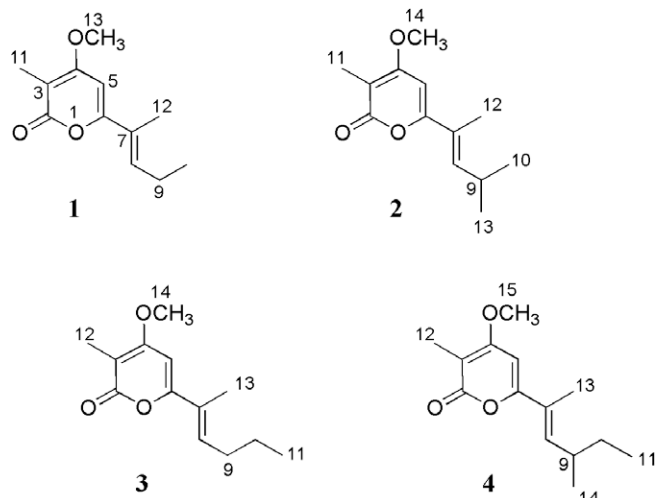
This superfamily contains three families: Polybranchiidae, Hermaeidae and Limapontiidae.¹⁷ Chemical studies conducted on several species belonging to Polybranchiidae and Limapontiidae families have shown that, with the exception of *Costasiella*

species,¹⁸ they contain α - and γ -pyrone polyketides de novo biosynthesised.¹⁹ The multifunctional biological roles of these molecules include an involvement in the chemical defence as well as in the regeneration of the detached appendages. The individuation of the true source of the carbon skeletons of pyrone-containing propionates has been object of several investigations. However, the de novo origin through a mixed acetate/propionate pathway has been suggested to be the most plausible hypothesis in molluscs.¹⁹ This suggestion has been very recently confirmed by incorporation of both propionate and acetate precursors labelled with stable isotopes in the Mediterranean sacoglossan *P. dendritica*.²⁰ To date, no chemical study was reported for molluscs belonging to the Hermaeidae family.

We describe here the finding of four α -pyrone polypropionates, named aplysiopsenes A–D (1–4), in the hermaeidean sacoglossan *Aplysiopsis formosa* Pruvot-Fol. Interestingly, these molecules display a shorter side chain with respect to related pyrone-polypropionates reported from the other limapontioidean sacoglossans.^{9–16}

Fourteen *A. formosa* individuals (20–25 mm average size) were collected along the coasts of São Jorge Island in the Azores (northern Atlantic Ocean), immediately frozen and then transferred to ICB in Naples. The frozen material was repeatedly extracted with acetone (50 ml \times 3) at room temperature and the aqueous residue, after evaporation of the organic solvent under reduced pressure, was partitioned between water and Et₂O. The Et₂O extract (11.7 mg) was chromatographed by silica gel column using a

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gradient eluent system from light petroleum to Et₂O. The fractions obtained with light petroleum ether/Et₂O 85:15 to 8:2 were further purified by analytical RP-HPLC (gradient MeOH/H₂O, from 60% to 100% MeOH, UV detector 210 nm) to yield in order of decreasing polarity aplysiopsene A (**1**, Rt 17.3), B (**2**, Rt 24.1), C (**3**, Rt 25.0) and D (**4**, Rt 31.4).

Aplysiopsene A (**1**)²¹ was isolated as a colourless oil and showed a sodiated-molecular peak at *m/z* 231.0998 in the HRESIMS spectrum (231.0997 calculated for C₁₂H₁₆O₃Na), which indicated the molecular formula C₁₂H₁₆O₃. An intense IR band at ν_{\max} 1678 cm⁻¹ and the presence of signals at δ 6.10 (1H, s, H-5), 3.90 (3H, s, H₃-13) and 1.94 (3H, s, H₃-11) in the ¹H NMR spectrum as well as at δ 165.9 (C-4), 164.4 (C-2), 160.1 (C-6) and 91.4 (C-5) in the ¹³C NMR spectrum (Table 1) suggested the presence of a 3-methyl-4-methoxy-6-substituted- α -pyrone system. The ¹H NMR spectrum of **1** showed additional signals at δ 6.61 (1H, t, *J* = 7.3, H-8), 2.24 (2H, quintet, *J* = 7.3, H₂-9), 1.88 (3H, br s, H₃-12) and 1.08 (3H, t, *J* = 7.3, H₃-10) that were attributed to a 2-pentenyl alkyl chain linked to C-6 of the α -pyrone ring as showed in structure **1**. The geometry of the double bond at C-7 was inferred as *E* by both the carbon value of C-12 (δ 12.2) and a NOE effect observed between H₃-12 and H₂-9. Comparison of the spectral data of **1** with those reported in the literature for similar polypropionates isolated from both sacoglossans¹⁴ and fungi²² further supported the structure proposed. In particular, aplysiopsene A was closely related to

the fungal metabolite nectriapyrone²³ differing from the latter in the presence of an additional CH₂ unit in the side chain.

The spectroscopic data of aplysiopsene B (**2**)²⁴ were very similar to those of compound **1**. The sodiated-molecular peak at *m/z* 245.1157 in the HRESIMS spectrum (245.1154 calculated for C₁₃H₁₈O₃Na) indicated the molecular formula C₁₃H₁₈O₃ with an additional CH₂ moiety with respect to **1**. Analysis of both ¹H and ¹³C NMR spectra of **2** clearly indicated the presence of the same α -pyrone moiety, whereas the terminal moiety of the alkyl chain was different. In fact, typical signals attributable to an isopropyl residue at δ 2.70 (1H, dq, *J* = 6.7, 9.5, H-9) and δ 1.05 (6H, d, *J* = 6.7, H₃-10 and H₃-13) were observed in the ¹H NMR spectrum of **2** in the place of the signals due to the terminal ethyl group in compound **1**. The carbon value of C-12 (δ 12.0) was consistent with the 7*E*-stereochemistry.

The molecular formula C₁₃H₁₈O₃ of aplysiopsene C (**3**)²⁵ which was deduced by the sodiated-molecular peak at *m/z* 245.1149 in the HRESIMS spectrum, was the same (245.1154 calculated for C₁₃H₁₈O₃Na) as that of compound **2**. Analysis of both proton and carbon spectra of **3** showed that in this molecule, the alkyl chain linked to the α -pyrone moiety contained a terminal *n*-propyl group. In fact, in the ¹H-¹H COSY spectrum of **3**, the olefinic proton resonating at δ 6.62 (1 H, t, *J* = 9.5, H-8) was correlated with a methylene at δ 2.20 (2 H, m, H₂-9) which in turn coupled with another methylene resonating at δ 1.50 (2 H, m, H₂-10) finally linked to a terminal methyl at δ 0.94 (3 H, t, *J* = 7.3, H₃-11). The remaining part of the molecule was confirmed to be the same as that of **1** and **2**.

Aplysiopsene D (**4**)²⁶ had the molecular formula C₁₄H₂₀O₃ as obtained by the HRESIMS sodiated peak at *m/z* 259.1311 (259.1310 calculated for C₁₄H₂₀O₃Na). The structural analogies with the co-occurring compounds **1**–**3** were evident by analysis of ¹H and ¹³C NMR spectra. As for the other molecules, the only difference was in the terminal part of the alkyl chain. In particular, the proton spectrum displayed in the high-field region two methyl signals at δ 1.02 (3H, d, *J* = 6.7, H₃-14) and δ 0.88 (3H, t, *J* = 7.5, H₃-11), a methine multiplet at δ 2.45 (1H, m, H-9), and a methylene signal at δ 1.35 (2H, m, H₂-10) according to the presence of a 2-butyl moiety linked to the olefinic proton H-8. Compound **4** showed strong structural similarities with fungal metabolite phenomenin B,²² being the corresponding 9,10 dihydroderivative.

All proton and carbon resonances of aplysiopsenes A–D were assigned by 2D NMR experiments.

Apparently, aplysiopsenes represent an additional example of marine polyketides with a mixed acetate/propionate pathway in

Table 1
¹H and ¹³C NMR data in CDCl₃ of aplysiopsenes (**1**–**4**)

C	1		2		3		4	
	δ ¹³ C ^a m ^b	δ ¹ H ^c m J (Hz)	δ ¹³ C ^a m ^b	δ ¹ H ^c m J (Hz)	δ ¹³ C ^a m ^b	δ ¹ H ^c m J (Hz)	δ ¹³ C ^a m ^b	δ ¹ H ^c m J (Hz)
2	164.4 s	—	164.4 s	—	165.2 s	—	164.4 s	—
3	102.2 s	—	102.2 s	—	102.2 s	—	102.1 s	—
4	165.9 s	—	165.7 s	—	165.7 s	—	165.4 s	—
5	91.4 d	6.10 s	91.5 d	6.10 s	91.6 d	6.10 s	91.5 d	6.10 s
6	160.1 s	—	159.5 s	—	160.1 s	—	160.2 s	—
7	125.8 s	—	123.8 s	—	126.0 s	—	124.6 s	—
8	136.8 d	6.61 t 7.3	141.9 d	6.46 d 9.5	135.4 d	6.62 t 7.3	141.1 d	6.42 d 9.5
9	21.6 t	2.24 quin 7.3	28.0 d	2.70 dq 6.7, 9.5	30.7 t	2.20 m	35.2 d	2.45 m
10	13.3 q	1.08 t 7.3	22.3 q	1.05 d 6.7	22.4 t	1.50 m	30.3 t	1.35 m
11	8.5 q	1.94 s	8.1 q	1.92 s	13.9 q	0.94 t 7.5	n.d.	0.88 t 7.5
12	12.2 q	1.88 s	12.0 q	1.88 s	8.6 q	1.94 s	8.0 q	1.94 s
13	55.9 q	3.90 s	22.3 q	1.05 d 6.7	12.4 q	1.89 s	10.8 q	1.90 s
14	—	—	55.9 q	3.90 s	56.1 q	3.90 s	19.8 q	1.02 d 6.1
15	—	—	—	—	—	—	55.4 q	3.91 s

^a Bruker 300 MHz spectrometer, chemical shifts (ppm) referred to CDCl₃ (δ 77.0).

^b Multiplicities deduced by DEPT experiment.

^c Bruker 400 and 600 MHz, assignments made by ¹H-¹H COSY, HSQC and HMBC (*J* = 10 Hz) experiments.

agreement with the structure features of secondary metabolites of limapontoidean sacoglossan species. However, this biosynthesis has to be confirmed by rigorous biosynthetic experiments. *A. formosa* is the first limapontoidean sacoglossan possessing only α -pyrone polyketides. In addition, the extra ring conjugation is limited to only one double bond. Probably *A. formosa* and the other studied polyketide-possessing limapontoideans should occupy in a hypothetical evolutionary scenario very near but distinct positions.

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- Compound **1**. 0.7 mg, 6.2%; UV λ_{max} (EtOH)/nm 228 (13,000), 321 (7500); HRESIMS m/z 231.0998 (231.0997 calculated for $\text{C}_{12}\text{H}_{16}\text{O}_3\text{Na}$); ^1H and ^{13}C NMR data are given in Table 1.
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- Compound **3**. 1.5 mg, 12.5%; IR $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 1681; UV λ_{max} (EtOH)/nm 228 (10,900), 321 (7000); HRESIMS m/z 245.1149 (245.1154 calculated for $\text{C}_{13}\text{H}_{18}\text{O}_3\text{Na}$); ^1H and ^{13}C NMR data are given in Table 1.
- Compound **4**. 0.4 mg, 3.4%, $[\alpha]_{\text{D}}^{25}$ -42.4 (c 0.04, CHCl_3); IR $\nu_{\text{max}}/\text{cm}^{-1}$ (film) 1681; UV λ_{max} (EtOH)/nm 228 (9800), 321 (7000); HRESIMS m/z 259.1311 (259.1310 calculated for $\text{C}_{14}\text{H}_{20}\text{O}_3\text{Na}$); ^1H and ^{13}C NMR data are given in Table 1.